

# UC Davis

## UC Davis Previously Published Works

### Title

Large-Scale Release of Campylobacter Draft Genomes: Resources for Food Safety and Public Health from the 100K Pathogen Genome Project.

### Permalink

<https://escholarship.org/uc/item/8mf4m9nk>

### Journal

Genome announcements, 5(1)

### ISSN

2169-8287

### Authors

Weis, Allison M  
Huang, Bihua C  
Storey, Dylan B  
et al.

### Publication Date

2017

### DOI

10.1128/genomea.00925-16

Peer reviewed



# Large-Scale Release of *Campylobacter* Draft Genomes: Resources for Food Safety and Public Health from the 100K Pathogen Genome Project

 Allison M. Weis,<sup>a,e</sup> Bihua C. Huang,<sup>a,e</sup> Dylan B. Storey,<sup>a,e\*</sup> Nguyet Kong,<sup>a,e</sup> Poyin Chen,<sup>a,e</sup> Narine Arabyan,<sup>a,e</sup> Brent Gilpin,<sup>b</sup> Carl Mason,<sup>c</sup> Andrea K. Townsend,<sup>d\*</sup> Woutrina A. Smith,<sup>a</sup> Barbara A. Byrne,<sup>a</sup> Conor C. Taff,<sup>d\*</sup>  Bart C. Weimer<sup>a,e</sup>

School of Veterinary Medicine, UC Davis, Davis, California, USA<sup>a</sup>; Institute of Environmental Science & Research Ltd., Christchurch, New Zealand<sup>b</sup>; Department of Enteric Diseases, Armed Forces Research Institute of Medical Sciences (AFRIMS), Bangkok, Thailand<sup>c</sup>; Department of Wildlife, Fisheries and Conservation Biology, UC Davis, Davis, California, USA<sup>d</sup>; 100K Pathogen Genome Project, UC Davis, Davis, California, USA<sup>e</sup>

**ABSTRACT** *Campylobacter* is a food-associated bacterium and a leading cause of foodborne illness worldwide, being associated with poultry in the food supply. This is the initial public release of 202 *Campylobacter* genome sequences as part of the 100K Pathogen Genome Project. These isolates represent global genomic diversity in the *Campylobacter* genus.

*Campylobacter* is the most common foodborne pathogen worldwide in humans and animals (1). Approximately 1.3 million people are infected in the United States yearly (2). Despite control efforts, outbreaks are increasing (2). Symptoms include fever, abdominal cramping, and bloody diarrhea, and in rare cases, infection leads to autoimmune disorders, such as Guillain-Barré syndrome (8). Infection is primarily from poultry but is also associated with domesticated livestock (3, 4) and wildlife (5–7).

The 100K Pathogen Genome Project (<http://www.100kgenomes.org>) is a large-scale sequencing effort for worldwide isolates with a genome repository at the 100K Project BioProject at the NCBI (PRJNA186441). This project included the three most commonly identified species of *Campylobacter*: *C. jejuni*, *C. lari*, and *C. coli*. For an in-depth review of *Campylobacter* molecular biology and pathogenesis, see the papers by Silva et al. (9) and Young et al. (10).

All *Campylobacter* isolates from the 100K Pathogen Genome Project were collected and banked in the laboratory of Bart Weimer (University of California, Davis, CA). Isolates were checked for purity and stored in liquid nitrogen (11). Genomic DNA (gDNA) was extracted from cultures grown on 5% blood agar plates (UC Davis, VetMed Biological Services) for one to two days, lysed (12), purified with a Qiagen QIAamp DNA minikit (catalog no. 51306), and analyzed on Agilent 2200 TapeStation system with the Genomic DNA ScreenTape assay for integrity of gDNA (13). After isolation, gDNA was fragmented using Diagenode Bioruptor next-generation sequencer (NGS) or Covaris E220 (14). Fragmented gDNA (1  $\mu$ g) was used for library construction with the Kapa high-throughput (HTP) library preparation kit (catalog no. KK8234; Kapa Biosystems, Boston, MA), using the Agilent Bravo NGS workstation (Santa Clara, CA). Fragmented double-strand gDNA molecules were end-repaired (5'), adenylated (3'), and ligated with double-stranded DNA (dsDNA) adapters, either NEXTflex-96 DNA barcode (Bioo Scientific, Austin, TX), and multiplexed up to 96 isolates, or using Integrated DNA Technologies, Inc. Weimer 384 TS-LT DNA barcodes that allowed multiplexing up to 384 genomes. The standard Kapa protocol with dual-SPRI size selection was used for 250-

Received 14 July 2016 Accepted 25 October 2016 Published 5 January 2017

**Citation** Weis AM, Huang BC, Storey DB, Kong N, Chen P, Arabyan N, Gilpin B, Mason C, Townsend AK, Smith WA, Byrne BA, Taff CC, Weimer BC. 2017. Large-scale release of *Campylobacter* draft genomes: resources for food safety and public health from the 100K Pathogen Genome Project. *Genome Announc* 5:e00925-16. <https://doi.org/10.1128/genomeA.00925-16>.

**Copyright** © 2017 Weis et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Bart C. Weimer, [bcweimer@ucdavis.edu](mailto:bcweimer@ucdavis.edu).

\* Present address: Dylan B. Storey, West Sacramento, California, USA; Andrea K. Townsend, Department of Biology, Hamilton College, Clinton, New York, USA; Conor C. Taff, Lab of Ornithology, Cornell University, Ithaca, New York, USA.

to 500-bp fragments. Library amplification was done for eight cycles using the Kapa HiFi HotStart ReadyMix, followed by a 1× SPRI bead cleanup step. Size distribution of libraries was confirmed using Agilent 2100 Bioanalyzer system with high-sensitivity DNA kit (15, 16), and indexed libraries were quantified with quantitative PCR (qPCR)-based Kapa library quantification kit (catalog no. KK4824) prior to pooling for sequencing either on the Illumina HiSeq 2000 with PE100 plus index read at BGI@UCD (Sacramento, CA) or PE150 on the Illumina HiSeq 3000 at the UC Davis Genome Center (Davis, CA). Paired-end reads were assembled using ABySS 1.5.2 using  $k = 64$  (17).

All sample sequences are publicly available on the NCBI's Sequence Read Archive (SRA) database (<http://www.ncbi.nlm.nih.gov/sra>), and genome assemblies can be found in NCBI GenBank. Here, the 100K Pathogen Genome Project has assembled 202 genomes from different isolates of *Campylobacter* identified as *C. jejuni* (167 genomes), *C. coli* (32 genomes), and *C. lari* (3 genomes) (Table 1).

**Accession number(s).** Sequences can be found in the 100K Project BioProject at the NCBI SRA BioProject and in the NCBI Genbank. Accession numbers are presented in Table 1.

## ACKNOWLEDGMENTS

We thank Whitney Ng and Kao Thao for their effort in isolate logistics and technical assistance and all of the collaborators to the 100K Pathogen Genome Project.

This project was funded by 100K Pathogen Genome Project, with initial funding from the FDA, and Agilent Technologies, to produce these sequences.

## REFERENCES

1. Coker AO, Isokpehi RD, Thomas BN, Amisu KO, Obi CL. 2002. Human campylobacteriosis in developing countries. *Emerg Infect Dis* 8:237–244. <https://doi.org/10.3201/eid0803.010233>.
2. CDC. 2012. Foodborne diseases Active Surveillance Network (FoodNet): FoodNet surveillance report for 2011 (final report). Centers for Disease Control and Prevention, Department of Health and Human Services, Atlanta, GA. [http://www.cdc.gov/foodnet/pdfs/2011\\_annual\\_report\\_508c.pdf](http://www.cdc.gov/foodnet/pdfs/2011_annual_report_508c.pdf).
3. Nielsen EM. 2002. Occurrence and strain diversity of thermophilic *campylobacters* in cattle of different age groups in dairy herds. *Lett Appl Microbiol* 35:85–89. <https://doi.org/10.1046/j.1472-765X.2002.01143.x>.
4. Jacobs-Reitsma W, Lyhs U, Wagenaar J. 2008. *Campylobacter* in the food supply, p 627–644. In Nachamkin I, Szymanski CM, Blaser MJ (ed), *Campylobacter*, 3rd ed. ASM Press, Washington, DC.
5. Weis AM, Miller WA, Byrne BA, Chouicha N, Boyce WM, Townsend AK. 2014. Prevalence and pathogenic potential of *Campylobacter* isolates from free-living, human-commensal American crows. *Appl Environ Microbiol* 80:1639–1644. <https://doi.org/10.1128/AEM.03393-13>.
6. Taff CC, Weis AM, Wheeler S, Hinton MG, Weimer BC, Barker CM, Jones M, Logsdon R, Smith WA, Boyce WM, Townsend AK. 2016. Influence of host ecology and behavior on *Campylobacter jejuni* prevalence and environmental contamination risk in a synanthropic wild bird. *Appl Environ Microbiol* 82:4811–4820. <https://doi.org/10.1128/AEM.01456-16>.
7. Gardner TJ, Fitzgerald C, Xavier C, Klein R, Pruckler J, Stroika S, McLaughlin JB. 2011. Outbreak of campylobacteriosis associated with consumption of raw peas. *Clin Infect Dis* 53:26–32. <https://doi.org/10.1093/cid/cir249>.
8. Altekruze SF, Stern NJ, Fields PI, Swerdlow DL. 1999. *Campylobacter jejuni*—an emerging foodborne pathogen. *Emerg Infect Dis* 5:28–35. <https://doi.org/10.3201/eid0501.990104>.
9. Silva J, Leite D, Fernandes M, Mena C, Gibbs PA, Teixeira P. 2011. *Campylobacter* spp. as a foodborne pathogen: a review. *Front Microbiol* 2:200. <https://doi.org/10.3389/fmicb.2011.00200>.
10. Young KT, Davis LM, Dirita VJ. 2007. *Campylobacter jejuni*: molecular biology and pathogenesis. *Nat Rev Microbiol* 5:665–679. <https://doi.org/10.1038/nrmicro1718>.
11. Kong N, Ng W, Lee V, Kelly L, Weimer BC. 2013. Production and analysis of high molecular weight genomic DNA for NGS pipelines using Agilent DNA extraction kit (p/n 200600). Application note. Agilent Technologies, Santa Clara, CA. <http://www.agilent.com/cs/library/applications/5991-3722EN.pdf>.
12. Jeannotte R, Lee E, Kong N, Ng W, Kelly L, Weimer BC. 2014. High-throughput analysis of foodborne bacterial genomic DNA using Agilent 2200 TapeStation and genomic DNA ScreenTape system. Application note. Agilent Technologies, Santa Clara, CA. <http://www.agilent.com/cs/library/applications/5991-4003EN.pdf>.
13. Kong N, Ng W, Cai L, Leonardo A, Kelly L, Weimer BC. 2014. Integrating the DNA integrity number (DIN) to assess genomic DNA (gDNA) quality control using the Agilent 2200 TapeStation system. Application note. Agilent Technologies, Santa Clara, CA. [http://hpst.cz/sites/default/files/attachments/integrating-dna-integrity-number-din-assess-genomic-dna-gdna-quality-control-using-agilent-2200\\_1.pdf](http://hpst.cz/sites/default/files/attachments/integrating-dna-integrity-number-din-assess-genomic-dna-gdna-quality-control-using-agilent-2200_1.pdf).
14. Jeannotte R, Lee E, Arabyan N, Kong N, Thao K, Huang BH, Kelly L, Weimer BC. 2014. Optimization of Covaris settings for shearing bacterial genomic DNA by focused ultrasonication and analysis using Agilent 2200 TapeStation. Application note. Agilent Technologies, Santa Clara, CA. <http://cn.agilent.com/cs/library/applications/5991-5075EN.pdf>.
15. Kong N, Ng W, Foutouhi A, Huang BH, Kelly L, Weimer BC. 2014. Quality control of high-throughput library construction pipeline for KAPA HTP library using an Agilent 2200 TapeStation. Application note. Agilent Technologies, Santa Clara, CA. <http://www.agilent.com/cs/library/applications/5991-5141EN.pdf>.
16. Kong N, Thao K, Huang C, Appel M, Lappin S, Knapp L, Kelly L, Weimer BC. 2014. Automated library construction using KAPA library preparation kits on the Agilent NGS workstation yields high-quality libraries for whole-genome sequencing on the Illumina platform. Application note. Agilent Technologies, Santa Clara, CA. <https://doi.org/10.13140/RG.2.1.2306.1203>.
17. Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJM, Birol I. 2009. ABySS: a parallel assembler for short read sequence data. *Genome Res* 19:1117–1123. <https://doi.org/10.1101/gr.089532.108>.

**TABLE 1** Species, isolate name, assembly data, and accession numbers of 202 *Campylobacter* genomes

GenBank accession no.	Isolate name	Species <sup>a</sup>	No. of contigs	Total genome size (bp)	No. of CDSs <sup>b</sup>
MJWB000000000	BCW_4453	<i>C. coli</i>	34	1,746,170	1,766
MJVZ000000000	BCW_4454	<i>C. coli</i>	46	1,983,591	2,062
MJWE000000000	BCW_4455	<i>C. coli</i>	43	1,872,803	1,937
MJWC000000000	BCW_4457	<i>C. coli</i>	37	1,834,436	1,893
MKAY000000000	BCW_4724	<i>C. coli</i>	31	1,711,767	1,724
MKCC000000000	BCW_5137	<i>C. coli</i>	25	1,725,970	1,742
MKAO000000000	BCW_5818	<i>C. coli</i>	48	1,837,036	1,907
MJYL000000000	BCW_5914	<i>C. coli</i>	53	1,819,402	1,890
MJWD000000000	BCW_5916	<i>C. coli</i>	32	1,828,621	1,894
MJWF000000000	BCW_5917	<i>C. coli</i>	37	1,823,469	1,893
MJYV000000000	BCW_5918	<i>C. coli</i>	40	1,964,242	2,043
MJWL000000000	BCW_6447	<i>C. coli</i>	32	1,861,852	1,912
MJWM000000000	BCW_6448	<i>C. coli</i>	31	1,857,061	1,912
MJWA000000000	BCW_6450	<i>C. coli</i>	48	1,983,760	2,061
MJYP000000000	BCW_6860	<i>C. coli</i>	48	1,731,589	1,766
MJZG000000000	BCW_6913	<i>C. coli</i>	41	1,812,371	1,882
MJZH000000000	BCW_6914	<i>C. coli</i>	43	1,827,738	1,884
MKEX000000000	BCW_6946	<i>C. coli</i>	46	1,828,827	1,893
MJZK000000000	BCW_6948	<i>C. coli</i>	35	1,740,538	1,767
MJZJ000000000	BCW_6949	<i>C. coli</i>	43	1,823,646	1,888
MJZL000000000	BCW_6950	<i>C. coli</i>	34	1,740,481	1,766
MJZM000000000	BCW_6951	<i>C. coli</i>	57	1,966,450	2,056
MJZP000000000	BCW_6955	<i>C. coli</i>	41	1,818,341	1,886
MJZR000000000	BCW_6957	<i>C. coli</i>	47	1,868,605	1,928
MJZS000000000	BCW_6958	<i>C. coli</i>	40	1,738,043	1,774
MJZU000000000	BCW_7432	<i>C. coli</i>	35	1,874,299	1,934
MJZV000000000	BCW_7433	<i>C. coli</i>	41	1,813,482	1,882
MJZW000000000	BCW_7434	<i>C. coli</i>	48	1,848,847	1,926
MJZX000000000	BCW_7435	<i>C. coli</i>	62	1,967,893	2,051
MJZY000000000	BCW_7437	<i>C. coli</i>	41	1,829,683	1,892
MKAA000000000	BCW_7460	<i>C. coli</i>	36	1,629,723	1,686
MKAB000000000	BCW_7692	<i>C. coli</i>	51	1,785,336	1,805
MEIB000000000	BCW_3781	<i>C. jejuni</i>	59	1,803,513	1,853
MJYK000000000	BCW_3782	<i>C. jejuni</i>	41	1,669,259	1,677
MEIC000000000	BCW_3784	<i>C. jejuni</i>	30	1,748,672	1,864
MJWO000000000	BCW_3785	<i>C. jejuni</i>	28	1,731,393	1,759
MJWP000000000	BCW_3786	<i>C. jejuni</i>	76	1,808,881	1,856
MJVF000000000	BCW_3787	<i>C. jejuni</i>	75	1,805,330	1,856
MJVG000000000	BCW_3788	<i>C. jejuni</i>	28	1,673,630	1,695
MJVH000000000	BCW_3789	<i>C. jejuni</i>	31	1,681,150	1,765
MJVI000000000	BCW_3790	<i>C. jejuni</i>	75	1,809,651	1,836
MJVJ000000000	BCW_3791	<i>C. jejuni</i>	141	2,006,566	2,034
MJVK000000000	BCW_3792	<i>C. jejuni</i>	74	1,809,576	1,833
MJVL000000000	BCW_3794	<i>C. jejuni</i>	63	1,759,193	1,775
MJVM000000000	BCW_3797	<i>C. jejuni</i>	31	1,699,040	1,676
MJVN000000000	BCW_3798	<i>C. jejuni</i>	53	1,744,518	1,751
MJVO000000000	BCW_3799	<i>C. jejuni</i>	55	1,768,431	1,783
MJVP000000000	BCW_3800	<i>C. jejuni</i>	70	1,832,983	1,840
MJVQ000000000	BCW_3802	<i>C. jejuni</i>	64	1,836,234	1,848
MJVR000000000	BCW_3803	<i>C. jejuni</i>	19	1,693,605	1,689
MKAL000000000	BCW_3804	<i>C. jejuni</i>	42	1,700,860	1,688
MJVS000000000	BCW_3805	<i>C. jejuni</i>	74	1,807,158	1,832
MJVT000000000	BCW_3807	<i>C. jejuni</i>	95	1,997,747	2,027
MJVU000000000	BCW_3810	<i>C. jejuni</i>	67	1,827,740	1,836
MJVV000000000	BCW_4216	<i>C. jejuni</i>	29	1,689,963	1,694
MKAM000000000	BCW_4218	<i>C. jejuni</i>	36	1,688,993	1,707
MJXR000000000	BCW_4219	<i>C. jejuni</i>	29	1,689,256	1,693
MJXS000000000	BCW_4220	<i>C. jejuni</i>	71	1,812,101	1,834
MJWI000000000	BCW_4221	<i>C. jejuni</i>	33	1,660,182	1,633
MJXT000000000	BCW_4222	<i>C. jejuni</i>	76	1,812,008	1,832
MJXU000000000	BCW_4223	<i>C. jejuni</i>	73	1,792,952	1,790

(Continued on next page)

TABLE 1 (Continued)

GenBank accession no.	Isolate name	Species <sup>a</sup>	No. of contigs	Total genome size (bp)	No. of CDSs <sup>b</sup>
MJWH000000000	BCW_4224	<i>C. jejuni</i>	29	1,680,989	1,689
MJXV000000000	BCW_4225	<i>C. jejuni</i>	72	1,808,664	1,827
MJXW000000000	BCW_4226	<i>C. jejuni</i>	29	1,693,524	1,706
MJXX000000000	BCW_4228	<i>C. jejuni</i>	74	1,776,092	1,777
MJXY000000000	BCW_4229	<i>C. jejuni</i>	76	1,777,005	1,776
MJWJ000000000	BCW_4230	<i>C. jejuni</i>	34	1,670,410	1,639
MJXZ000000000	BCW_4231	<i>C. jejuni</i>	66	1,833,811	1,842
MKAW000000000	BCW_4317	<i>C. jejuni</i>	87	1,729,089	1,758
MKAQ000000000	BCW_4319	<i>C. jejuni</i>	18	1,662,736	1,639
MJYA000000000	BCW_4321	<i>C. jejuni</i>	67	1,840,105	1,882
MJYB000000000	BCW_4322	<i>C. jejuni</i>	67	1,806,657	1,837
MJYC000000000	BCW_4323	<i>C. jejuni</i>	88	1,736,882	1,756
MKET000000000	BCW_4324	<i>C. jejuni</i>	56	1,804,461	1,856
MJYD000000000	BCW_4325	<i>C. jejuni</i>	88	1,733,375	1,755
MJYE000000000	BCW_4326	<i>C. jejuni</i>	90	1,732,137	1,750
MKAN000000000	BCW_4328	<i>C. jejuni</i>	63	1,854,445	1,889
MJYF000000000	BCW_4332	<i>C. jejuni</i>	70	1,739,132	1,764
MJYG000000000	BCW_4333	<i>C. jejuni</i>	81	1,874,835	1,927
MJYH000000000	BCW_4335	<i>C. jejuni</i>	91	1,725,603	1,756
MJYI000000000	BCW_4337	<i>C. jejuni</i>	68	1,806,323	1,836
MJYJ000000000	BCW_4338	<i>C. jejuni</i>	66	1,807,200	1,840
MKAH000000000	BCW_4341	<i>C. jejuni</i>	55	1,773,112	1,773
MJWG000000000	BCW_4452	<i>C. jejuni</i>	25	1,800,178	1,850
MKAI000000000	BCW_4456	<i>C. jejuni</i>	30	1,724,619	1,726
MJWN000000000	BCW_4459	<i>C. jejuni</i>	55	1,942,729	2,031
MKAJ000000000	BCW_4460	<i>C. jejuni</i>	25	1,601,734	1,576
MKAK000000000	BCW_4461	<i>C. jejuni</i>	22	1,604,851	1,577
MKAZ000000000	BCW_4727	<i>C. jejuni</i>	13	1,678,985	1,684
MKBA000000000	BCW_4728	<i>C. jejuni</i>	41	1,712,512	1,743
MKBB000000000	BCW_4731	<i>C. jejuni</i>	32	1,698,127	1,731
MKBC000000000	BCW_4734	<i>C. jejuni</i>	34	1,684,111	1,732
MKBD000000000	BCW_4735	<i>C. jejuni</i>	35	1,837,130	1,892
MKBE000000000	BCW_4737	<i>C. jejuni</i>	36	1,642,302	1,692
MKBF000000000	BCW_4738	<i>C. jejuni</i>	40	1,748,256	1,788
MKBG000000000	BCW_4741	<i>C. jejuni</i>	172	1,819,989	1,970
MKBH000000000	BCW_4743	<i>C. jejuni</i>	18	1,644,315	1,648
MKBI000000000	BCW_4744	<i>C. jejuni</i>	61	1,654,980	1,715
MKBL000000000	BCW_4747	<i>C. jejuni</i>	22	1,718,223	1,720
MKBJ000000000	BCW_4748	<i>C. jejuni</i>	49	1,747,262	1,782
MKBK000000000	BCW_4749	<i>C. jejuni</i>	48	1,704,441	1,757
MKBM000000000	BCW_4753	<i>C. jejuni</i>	60	1,661,715	1,720
MKBN000000000	BCW_4755	<i>C. jejuni</i>	27	1,687,997	1,681
MKBO000000000	BCW_4757	<i>C. jejuni</i>	51	1,645,272	1,708
MKBP000000000	BCW_5121	<i>C. jejuni</i>	52	1,891,211	1,916
MKBQ000000000	BCW_5122	<i>C. jejuni</i>	94	1,846,320	1,949
MKBR000000000	BCW_5123	<i>C. jejuni</i>	60	1,722,506	1,797
MKBS000000000	BCW_5124	<i>C. jejuni</i>	66	1,673,066	1,727
MKBT000000000	BCW_5125	<i>C. jejuni</i>	49	1,774,518	1,808
MKBU000000000	BCW_5126	<i>C. jejuni</i>	69	1,815,843	1,883
MKBV000000000	BCW_5128	<i>C. jejuni</i>	116	1,797,324	1,868
MKBW000000000	BCW_5129	<i>C. jejuni</i>	58	1,703,448	1,777
MKBX000000000	BCW_5131	<i>C. jejuni</i>	120	1,775,968	1,880
MKBY000000000	BCW_5132	<i>C. jejuni</i>	38	1,694,325	1,735
MKBZ000000000	BCW_5133	<i>C. jejuni</i>	140	1,858,971	1,987
MKCA000000000	BCW_5135	<i>C. jejuni</i>	105	1,776,102	1,902
MKCB000000000	BCW_5136	<i>C. jejuni</i>	52	1,780,266	1,796
MKCD000000000	BCW_5140	<i>C. jejuni</i>	42	1,691,709	1,726
MKCE000000000	BCW_5141	<i>C. jejuni</i>	73	1,701,529	1,760
MKCF000000000	BCW_5143	<i>C. jejuni</i>	95	1,795,032	1,848
MKCG000000000	BCW_5144	<i>C. jejuni</i>	95	1,787,787	1,880
MKCH000000000	BCW_5145	<i>C. jejuni</i>	65	1,752,372	1,798
MKCI000000000	BCW_5146	<i>C. jejuni</i>	60	1,764,810	1,832

(Continued on next page)

TABLE 1 (Continued)

GenBank accession no.	Isolate name	Species <sup>a</sup>	No. of contigs	Total genome size (bp)	No. of CDSs <sup>b</sup>
MKCJ000000000	BCW_5147	<i>C. jejuni</i>	130	1,721,583	1,824
MKCK000000000	BCW_5148	<i>C. jejuni</i>	42	1,670,278	1,703
MKCL000000000	BCW_5150	<i>C. jejuni</i>	98	1,794,347	1,876
MKCM000000000	BCW_5151	<i>C. jejuni</i>	53	1,683,276	1,727
MKEY000000000	BCW_5152	<i>C. jejuni</i>	41	1,685,669	1,721
MKEZ000000000	BCW_5154	<i>C. jejuni</i>	65	1,741,318	1,809
MKFA000000000	BCW_5155	<i>C. jejuni</i>	67	1,722,407	1,807
MKFB000000000	BCW_5156	<i>C. jejuni</i>	85	1,817,799	1,814
MKFC000000000	BCW_5157	<i>C. jejuni</i>	88	1,810,646	1,803
MKFD000000000	BCW_5158	<i>C. jejuni</i>	26	1,643,431	1,670
MKFE000000000	BCW_5159	<i>C. jejuni</i>	40	1,633,952	1,655
MKFF000000000	BCW_5160	<i>C. jejuni</i>	54	1,824,739	1,887
MKHS000000000	BCW_5161	<i>C. jejuni</i>	56	1,770,897	1,786
MKHT000000000	BCW_5162	<i>C. jejuni</i>	34	1,771,963	1,791
MKHU000000000	BCW_5166	<i>C. jejuni</i>	47	1,880,441	1,915
MKHV000000000	BCW_5167	<i>C. jejuni</i>	32	1,715,657	1,746
MKHW000000000	BCW_5170	<i>C. jejuni</i>	68	1,803,925	1,863
MKHX000000000	BCW_5171	<i>C. jejuni</i>	117	1,764,157	1,809
MKHY000000000	BCW_5172	<i>C. jejuni</i>	64	1,695,537	1,706
MKHZ000000000	BCW_5174	<i>C. jejuni</i>	98	1,757,175	1,779
MKE500000000	BCW_5913	<i>C. jejuni</i>	80	1,829,124	1,791
MJXA000000000	BCW_6451	<i>C. jejuni</i>	23	1,658,447	1,671
MJXB000000000	BCW_6452	<i>C. jejuni</i>	25	1,657,388	1,670
MJXC000000000	BCW_6453	<i>C. jejuni</i>	73	1,870,366	1,907
MJXD000000000	BCW_6454	<i>C. jejuni</i>	78	1,839,000	1,885
MJXE000000000	BCW_6456	<i>C. jejuni</i>	47	1,738,922	1,802
MJXF000000000	BCW_6457	<i>C. jejuni</i>	66	1,820,178	1,841
MJXG000000000	BCW_6458	<i>C. jejuni</i>	48	1,770,748	1,779
MJXH000000000	BCW_6459	<i>C. jejuni</i>	91	1,873,732	1,895
MJXI000000000	BCW_6460	<i>C. jejuni</i>	42	1,743,085	1,756
MJXJ000000000	BCW_6461	<i>C. jejuni</i>	28	1,724,268	1,755
MJXK000000000	BCW_6462	<i>C. jejuni</i>	45	1,762,804	1,774
MJXL000000000	BCW_6463	<i>C. jejuni</i>	80	1,810,501	1,847
MJXM000000000	BCW_6464	<i>C. jejuni</i>	89	1,874,875	1,906
MJXN000000000	BCW_6465	<i>C. jejuni</i>	113	1,898,179	1,918
MJXO000000000	BCW_6466	<i>C. jejuni</i>	75	1,856,101	1,908
MJXP000000000	BCW_6467	<i>C. jejuni</i>	52	1,706,044	1,706
MJXQ000000000	BCW_6468	<i>C. jejuni</i>	44	1,745,966	1,746
MJYM000000000	BCW_6475	<i>C. jejuni</i>	133	1,715,549	1,732
MJYN000000000	BCW_6476	<i>C. jejuni</i>	49	1,750,597	1,751
MJWQ000000000	BCW_6871	<i>C. jejuni</i>	83	1,838,932	1,893
MJWR000000000	BCW_6872	<i>C. jejuni</i>	38	1,727,268	1,728
MJWS000000000	BCW_6873	<i>C. jejuni</i>	114	1,864,402	1,881
MJWT000000000	BCW_6874	<i>C. jejuni</i>	116	1,864,348	1,871
MJWU000000000	BCW_6875	<i>C. jejuni</i>	40	1,737,054	1,756
MJWV000000000	BCW_6876	<i>C. jejuni</i>	39	1,798,356	1,914
MJWW000000000	BCW_6877	<i>C. jejuni</i>	65	1,866,607	1,905
MJWX000000000	BCW_6878	<i>C. jejuni</i>	114	1,862,090	1,875
MJWY000000000	BCW_6879	<i>C. jejuni</i>	31	1,633,912	1,684
MJYO000000000	BCW_6880	<i>C. jejuni</i>	40	1,684,005	1,685
MJYQ000000000	BCW_6881	<i>C. jejuni</i>	90	1,789,448	1,773
MJYR000000000	BCW_6882	<i>C. jejuni</i>	85	1,886,467	1,891
MJYS000000000	BCW_6883	<i>C. jejuni</i>	82	1,870,773	1,908
MKEW000000000	BCW_6884	<i>C. jejuni</i>	111	1,874,853	1,925
MJYT000000000	BCW_6885	<i>C. jejuni</i>	85	1,745,151	1,777
MJYU000000000	BCW_6886	<i>C. jejuni</i>	114	1,737,339	1,758
MKEV000000000	BCW_6887	<i>C. jejuni</i>	141	1,802,256	1,801
MJYV000000000	BCW_6888	<i>C. jejuni</i>	81	1,676,843	1,691
MJWZ000000000	BCW_6889	<i>C. jejuni</i>	81	1,944,460	2,003
MJYW000000000	BCW_6891	<i>C. jejuni</i>	83	1,751,933	1,778
MJYX000000000	BCW_6893	<i>C. jejuni</i>	57	1,770,259	1,778
MJYY000000000	BCW_6896	<i>C. jejuni</i>	120	1,625,057	1,606

(Continued on next page)

**TABLE 1** (Continued)

GenBank accession no.	Isolate name	Species <sup>a</sup>	No. of contigs	Total genome size (bp)	No. of CDSs <sup>b</sup>
MJYZ000000000	BCW_6897	<i>C. jejuni</i>	27	1,713,241	1,717
MJZA000000000	BCW_6898	<i>C. jejuni</i>	36	1,668,807	1,665
MJZB000000000	BCW_6899	<i>C. jejuni</i>	35	1,674,680	1,665
MJZC000000000	BCW_6900	<i>C. jejuni</i>	60	1,839,405	1,838
MJZD000000000	BCW_6901	<i>C. jejuni</i>	172	1,671,632	1,678
MKEU000000000	BCW_6902	<i>C. jejuni</i>	37	1,706,600	1,714
MJZE000000000	BCW_6904	<i>C. jejuni</i>	71	1,719,544	1,739
MKAP000000000	BCW_6907	<i>C. jejuni</i>	55	1,829,263	1,849
MJZF000000000	BCW_6910	<i>C. jejuni</i>	47	1,817,981	1,839
MJZN000000000	BCW_6953	<i>C. jejuni</i>	40	1,700,213	1,723
MJZO000000000	BCW_6954	<i>C. jejuni</i>	81	1,881,118	1,881
MJZQ000000000	BCW_6956	<i>C. jejuni</i>	57	1,783,446	1,795
MJZT000000000	BCW_6959	<i>C. jejuni</i>	48	1,699,337	1,726
MJZZ000000000	BCW_7438	<i>C. jejuni</i>	73	1,950,893	2,009
MJVX000000000	BCW_3783	<i>C. lari</i>	23	1,493,439	1,495
MJVV000000000	BCW_3793	<i>C. lari</i>	25	1,492,968	1,492
MJWK000000000	BCW_4217	<i>C. lari</i>	21	1,491,293	1,492

<sup>a</sup>The average number of contigs, genome size, and number of coding sequences were 41.4, 1,828,002.30 bp, and 1,884 for *C. coli*; 63.8, 1,764,345.40 bp, and 1,792 for *C. jejuni*; and 23, 1,492,566.50 bp, and 1,493 for *C. lari*, respectively.

<sup>b</sup>CDSs, coding sequences.